

POSTER PRESENTATION

Open Access

Evaluation of the impact of matrix stiffness on encapsulated HepaRG spheroids

Sofia P Rebelo^{1,2}, Marta Estrada^{1,2}, Rita Costa^{1,2}, Christophe Chesné³, Catarina Brito^{1,2}, Paula M Alves^{1,2*}

From 23rd European Society for Animal Cell Technology (ESACT) Meeting: Better Cells for Better Health
Lille, France. 23-26 June 2013

Background

The drug development process is widely hampered by the lack of human models that recapitulate liver functionality and efficiently predict toxicity of new chemical compounds. Moreover, liver failure is a global medical problem, with transplantation being the only effective treatment currently available. The bipotent liver progenitor cell line HepaRG can be differentiated into cholangiocyte and hepatocyte-like cells that express major functions of mature hepatocytes, representing a valuable tool to model hepatic function [1]. Current two-dimensional (2D) protocols for the differentiation into mature hepatocyte-like cells fail to recapitulate the complex cell-cell interactions, which are crucial for maintaining polarity and inherent mature hepatic functionality. Herein, we present a three-dimensional (3D) strategy for the culture of HepaRG cells based on the encapsulation of aggregates. The effect of matrix stiffness on expansion and differentiation was evaluated through encapsulation with different concentrations of alginate (1.1% and 2%). Further characterization of the hepatic features will reveal the extent of the hepatic functionality of the generated spheroids.

Materials and methods

HepaRG cells were routinely propagated in static conditions as previously described [2]. Briefly, culture medium Williams E was supplemented with 1% (v/v) Glutamax, 1% (v/v) pen/strep, 5 µg/ml insulin and 50 µM hydrocortisone hemisuccinate and 10% (v/v) FBS and cultures were maintained at 37 °C, 5% CO₂. Spinner vessels with ball impeller (Wheaton) were inoculated with inoculums ranging from 5 to 8 × 10⁵ cell/mL and

an agitation ranging from 35 to 45 rpm to attain the desired aggregation conditions. Aggregate size was determined by measuring Ferret's diameter using the Image J software (NIH). After 3 days of aggregation, spheroids were encapsulated in 1.1% and 2% (w/v) of Ultra Pure MVG alginate (UP MVG NovaMatrix, Pronova Biomedical) in NaCl 0.9% (w/v) solution. Encapsulation was performed in an electrostatically driven microencapsulation unit VarV1 (Nisco) and cultures were maintained for 14 days in stirred culture conditions. Viability was determined by the double stain viability test - alginate beads were collected from stirred cultures, incubated with fluorescein diacetate (10 µg/mL) and TO-PRO3® (1 µM) and observed on a fluorescence microscope (Leica DMI6000) - and by the Trypan blue exclusion method - alginate beads were dissociated with a solution of Sodium citrate 50 mM, Sodium chloride 104 mM and spheroids were dissociated by incubation with Trypsin 0.05%-EDTA (Gibco) and counted trypan blue exclusion dye. For characterization of the cultures, encapsulated spheroids were fixed as previously described [3] and incubated with phalloidin and prolong gold with DAPI and images were acquired in a confocal microscope (Andor spinning disk).

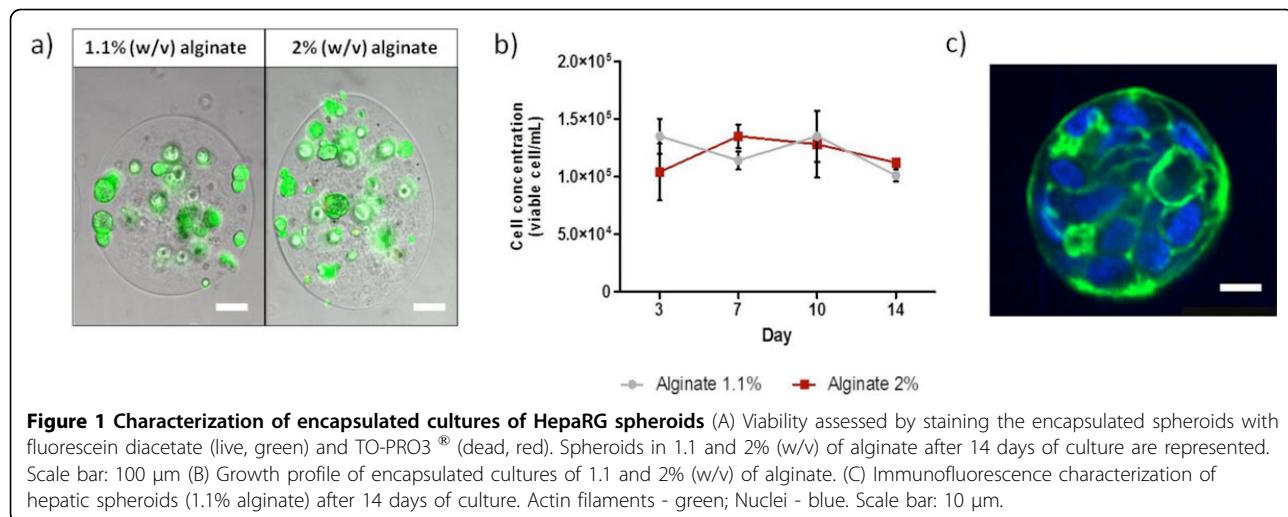
Results

In 2D cultures, HepaRG cells proliferate until confluence is reached and the cell-cell interactions established associated with the spatial constriction are postulated to trigger the differentiation program and maintain the differentiated state [1,4]. Moreover, the mechanochemical environment has been previously shown to strongly influence the liver-specific functions [5]. Thus, it was hypothesized that the microenvironment created by encapsulation of spheroids with an inert biomaterial with different stiffness levels, would promote differential behavior of the spheroids, towards differentiation or

* Correspondence: marques@itqb.unl.pt

¹iBET, Instituto de Biologia Experimental e Tecnológica, 2780-901 Oeiras, Portugal

Full list of author information is available at the end of the article



proliferation. Alginate concentrations of 1.1 and 2% (w/v) were used, given the 10 fold difference in stiffness, measured by the elastic modulus [6]. Both viability and the growth profile were monitored throughout culture time.

In both culture conditions, the viability was maintained above 85%, showing that the alginate concentration does not affect diffusion of nutrients or oxygen to supply effectively the cell spheroids (Figure 1 A). Moreover, it was observed that the growth profile was comparable for the two cultures, with growth arrest after aggregation and no proliferation occurring either in both alginate concentrations (Figure 1 B). This suggests that the differentiation program is triggered either in softer and stiffer microenvironments, being 1.1% alginate concentration sufficient to initiate the process.

The structural organization of the cell spheroids in both stiffness environments was characterized by the arrangement of actin filaments, which is associated to the tight junctions in highly polarized epithelial cells. As shown in Figure 1 C, the cells are disposed in a highly polarized manner, without necrotic centres.

Conclusions

In the current work, the encapsulation of liver spheroids with different stiffness conditions was evaluated as a strategy to culture HepaRG cells. It was observed that the encapsulation with different alginate concentrations is compatible with maintenance of highly viable cultures of liver spheroids, with growth arrest and cell polarization promoted by spatial constriction and the enhanced cell-cell interactions in 3D.

Acknowledgements

This work was supported by PTDC/EBB-BIO/112786/2009 and SFRH/BD/70264/2010 FCT, Portugal.

Authors' details

¹iBET, Instituto de Biologia Experimental e Tecnológica, 2780-901 Oeiras, Portugal. ²Instituto de Tecnologia Química e Biológica, Universidade Nova de Lisboa, 2780-157 Oeiras, Portugal. ³Biopredic International, Rennes, France (C. C., R.L., S.C.).

Published: 4 December 2013

References

1. Guillouzo A, Corlu A, Aninat C, Glaise D, Morel F, Guguen-Guillouzo C: The human hepatoma HepaRG cells: a highly differentiated model for studies of liver metabolism and toxicity of xenobiotics. *Chem Biol Interact* 2007, 168:66-73.
2. Grignon P, Rumin S, Urban S, Le Seyec J, Glaise D, Cannie I, Guyomard C, Lucas J, Trepo C, Guguen-Guillouzo C: Infection of a human hepatoma cell line by hepatitis B virus. *Proc Natl Acad Sci USA* 2002, 99:15655-15660.
3. Tostes RM, Leite SB, Serra M, Jensen J, BJORQUIST P, Carrondo MJ, Brito C, Alves PM: Human liver cell spheroids in extended perfusion bioreactor culture for repeated-dose drug testing. *Hepatology* 2012, 55:1227-1236.
4. Cerec V, Glaise D, Garnier D, Morosan S, Turlin B, Drenou B, Grignon P, Kremsdorff D, Guguen-Guillouzo C, Corlu A: Transdifferentiation of hepatocyte-like cells from the human hepatoma HepaRG cell line through bipotent progenitor. *Hepatology* 2007, 45:957-967.
5. Semler EJ, Ranucci CS, Moghe PV: Mechanochemical manipulation of hepatocyte aggregation can selectively induce or repress liver-specific function. *Biotechnol Bioeng* 2000, 69:359-369.
6. Martinsen A, Skjak-Braek G, Smidsrød O: Alginate as immobilization material: I. Correlation between chemical and physical properties of alginate gel beads. *Biotechnol Bioeng* 1989, 33:79-89.

doi:10.1186/1753-6561-7-S6-P77

Cite this article as: Rebelo et al.: Evaluation of the impact of matrix stiffness on encapsulated HepaRG spheroids. *BMC Proceedings* 2013 7 (Suppl 6):P77.